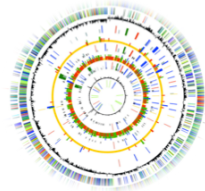




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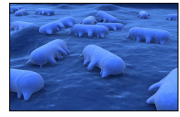
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Antimicrobial resistance- a global *One Health* challenge

Professor Séamus Fanning
UCD-Centre for Food Safety,
School of Public Health, Physiotherapy & Sports Science,
University College Dublin, Ireland.



Hosted by Professor Jean-Yves Maillard
Cardiff University, Wales



www.webbertraining.com February 18, 2020

Lecture overview -

- review of the discovery of antibiotics/antimicrobial compounds
- in brief, discuss the use of antimicrobial agents in a *One Health* context
- evaluate how resistance to antibiotics/antimicrobial compounds emerges and becomes disseminated
- examples of transmission of AMR genotypes
 - multi-drug resistant (MDR) *Salmonella* Indiana in poultry
 - detection of *mcr-1* and its variants
 - identification of *tet(X4)*-encoding resistance to tigecycline in *E. coli* cultured from food-producing animals
- analyse the metagenomics of antimicrobial resistance (AMR)

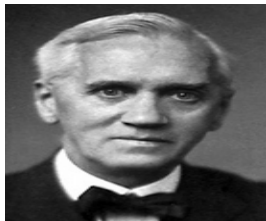
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Discovery of antibiotics/antimicrobial compounds -

3



Sir Alexander Fleming
Prize share: 1/3



Ernst Boris Chain
Prize share: 1/3

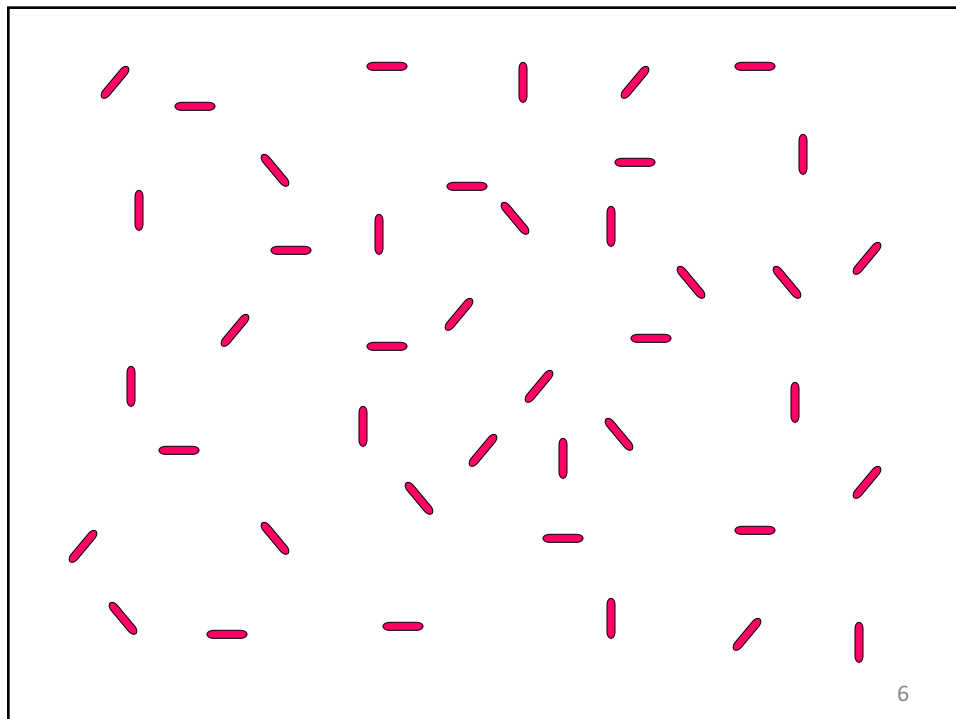
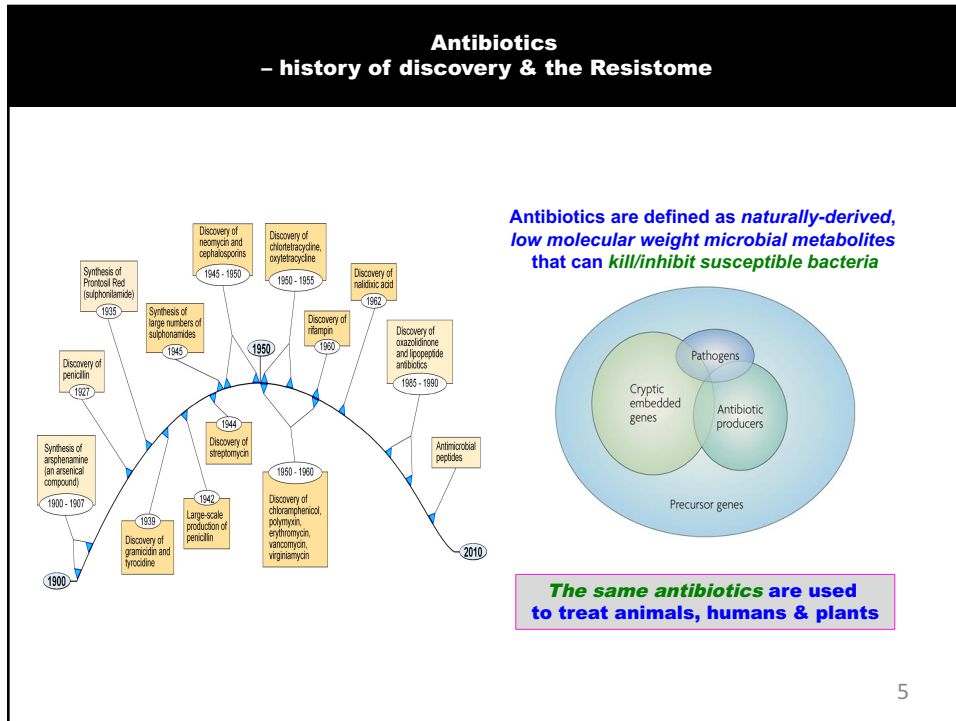


Sir Howard Walter Florey
Prize share: 1/3

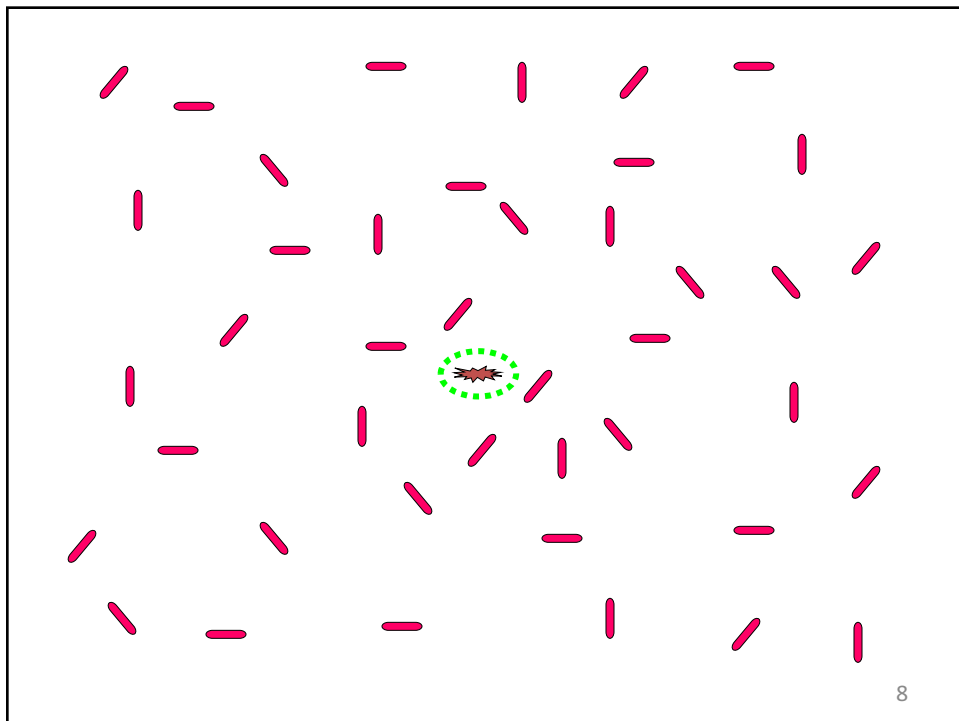
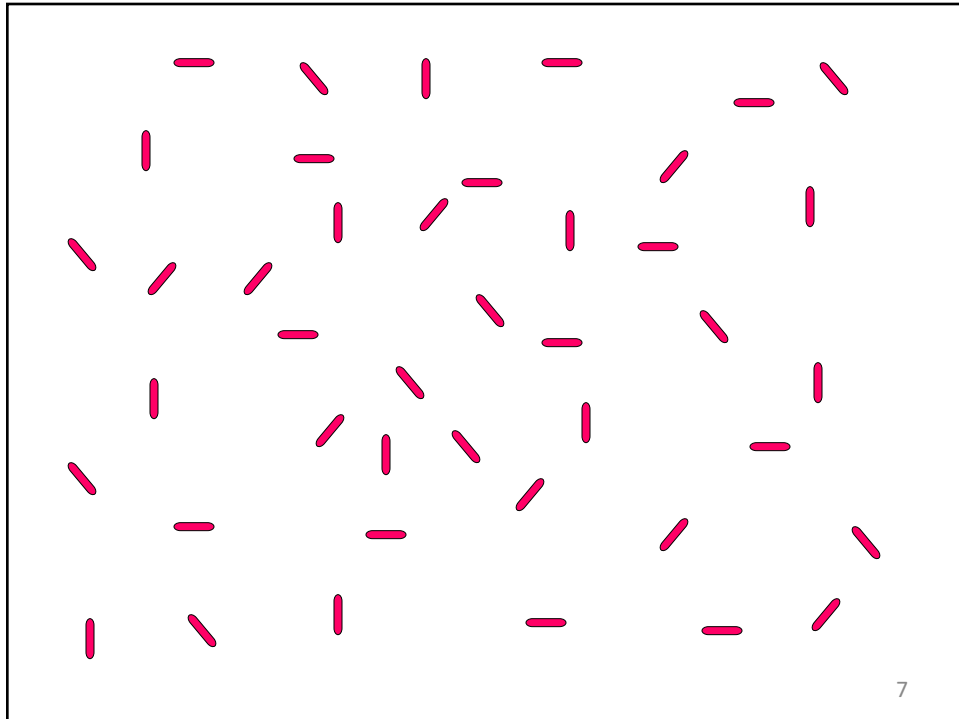
- **Antibiotics have revolutionized modern medicine** and enables physicians to undertake new treatments, such as surgery; organ transplantation; cancer chemotherapy among others
- these drugs are **among the most commonly prescribed medicines**, with some 70 billion clinical doses administered in 2010
- **antibiotics are naturally occurring molecules and can be purified from many soil bacteria, with many of these compounds being detected in pristine environments, that pre-date human use**
- **it is not surprising to note that defense or resistance to these molecules is also detectable**
- DNA sequencing of ancient permafrost sediment samples, detected genes that conferred resistance to several antimicrobial classes

4

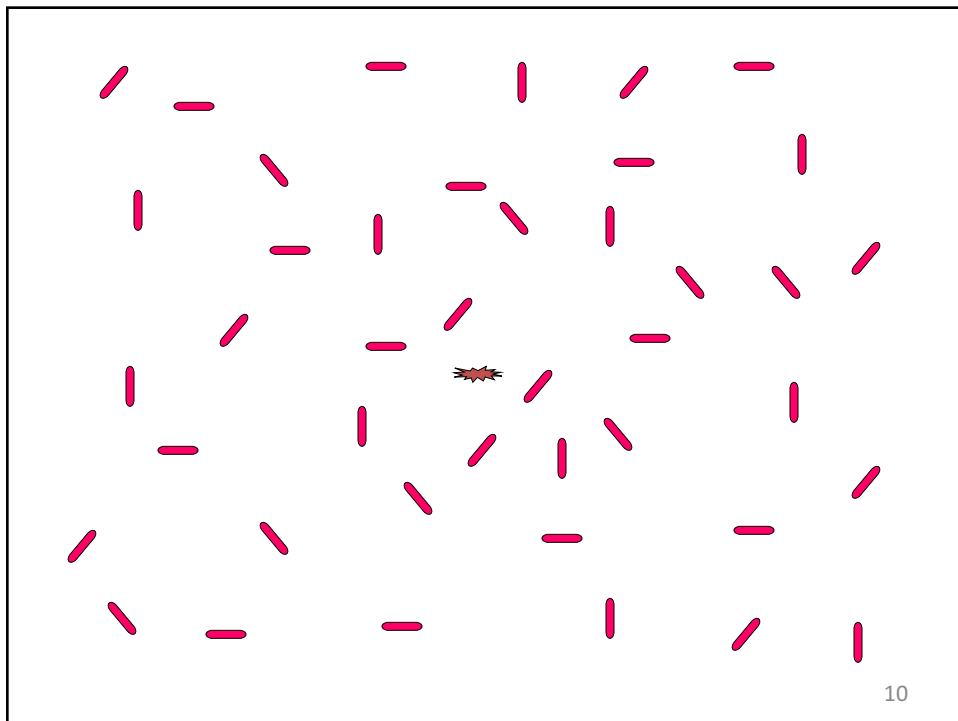
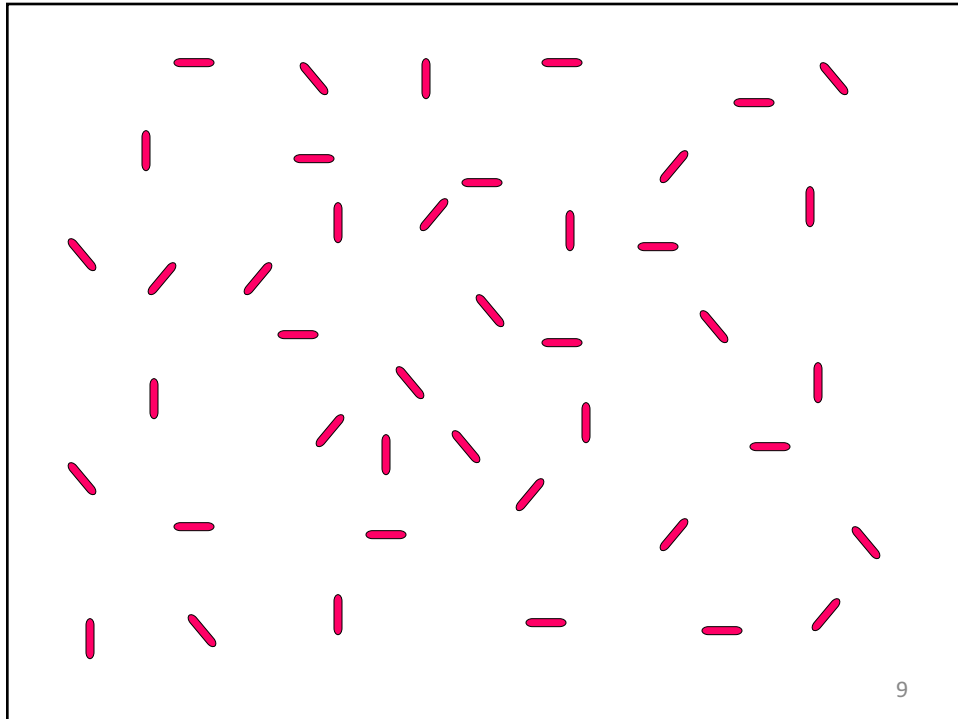
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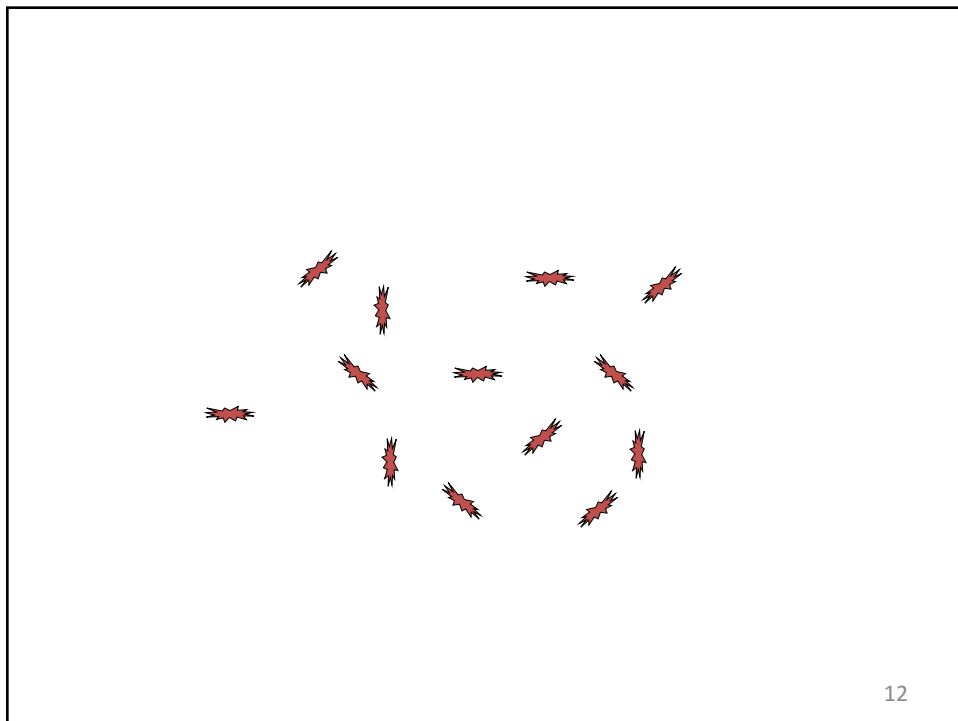
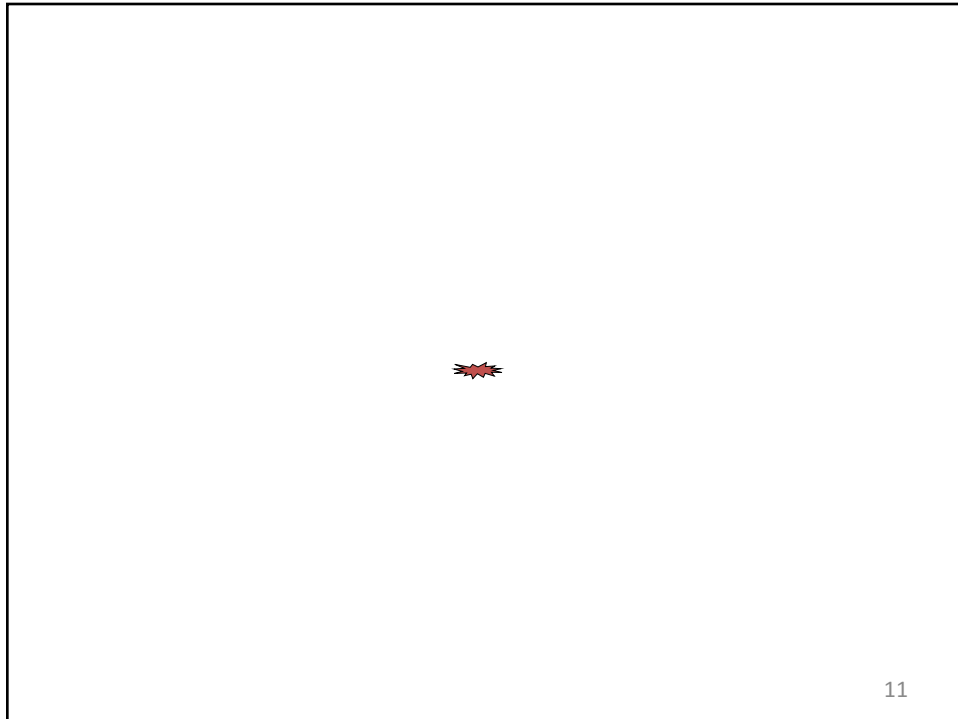
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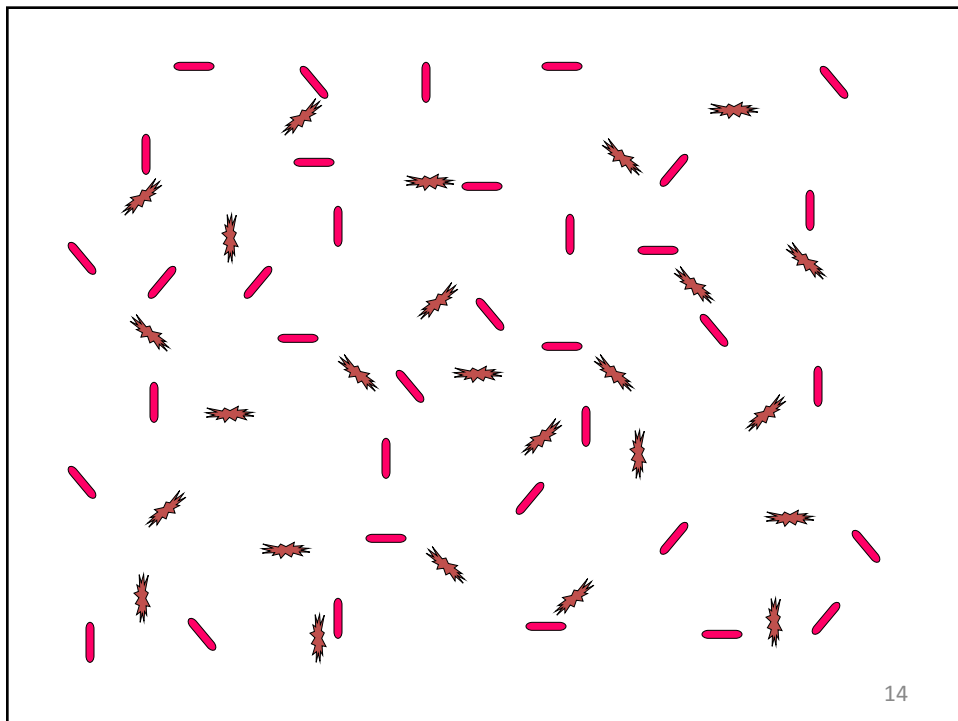
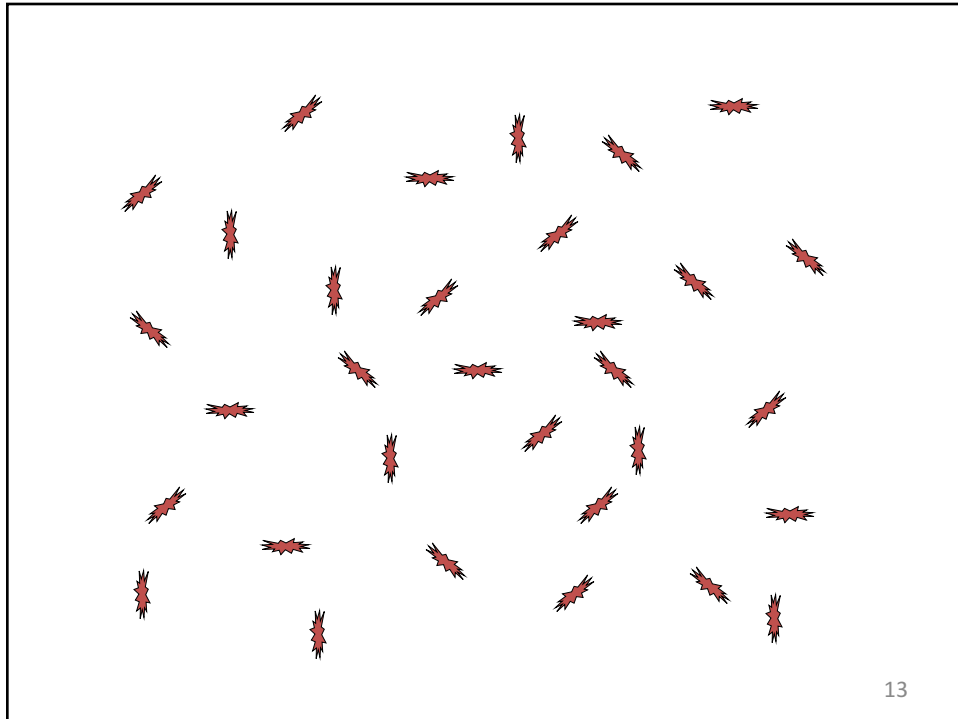
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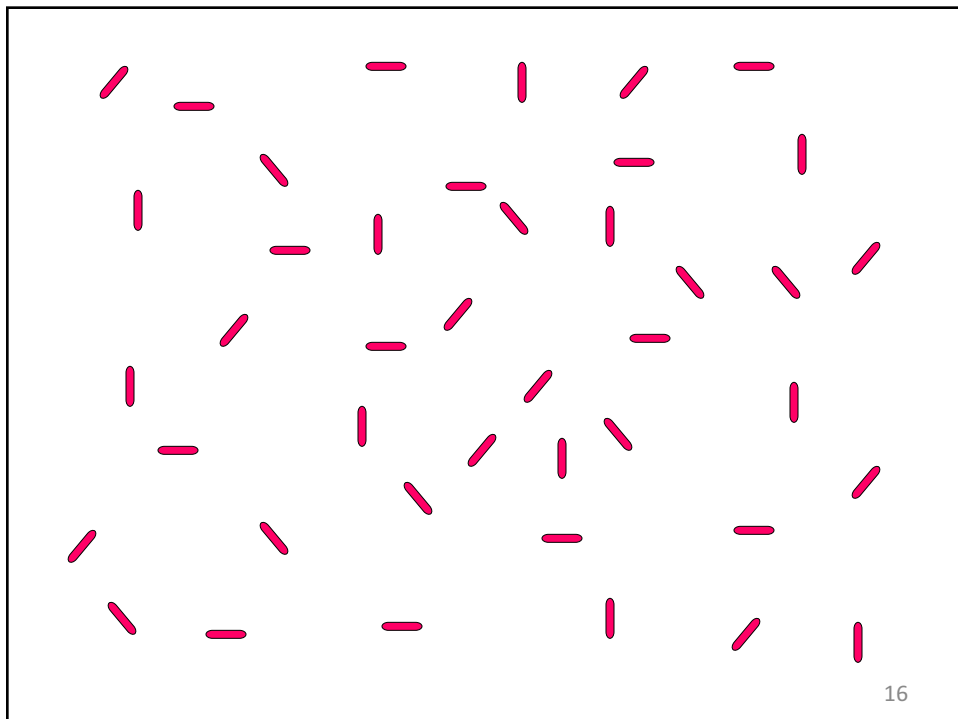
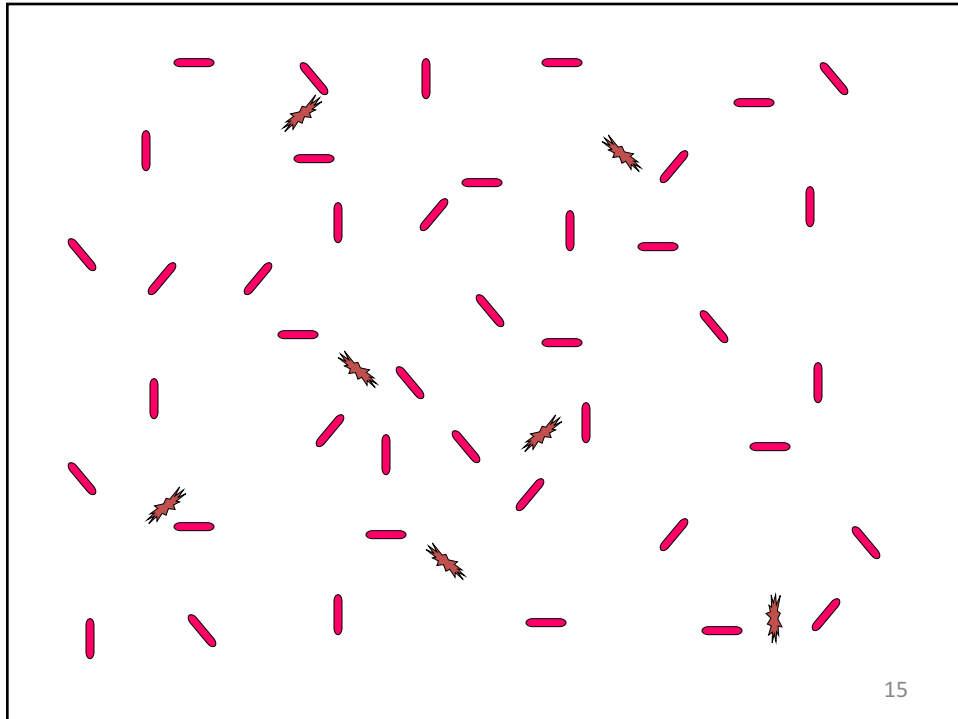


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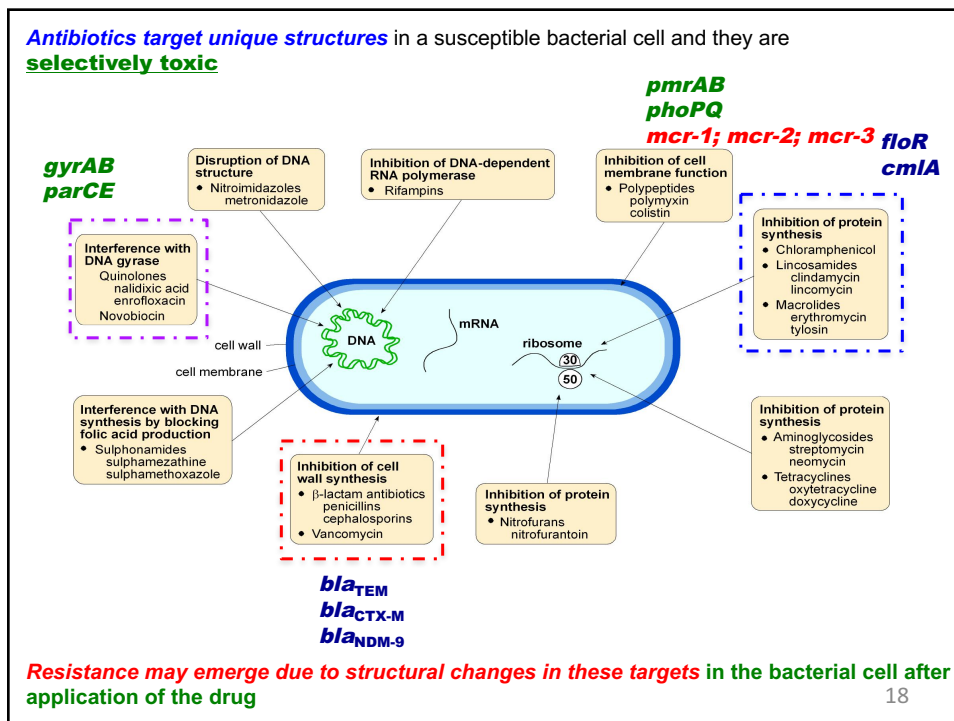
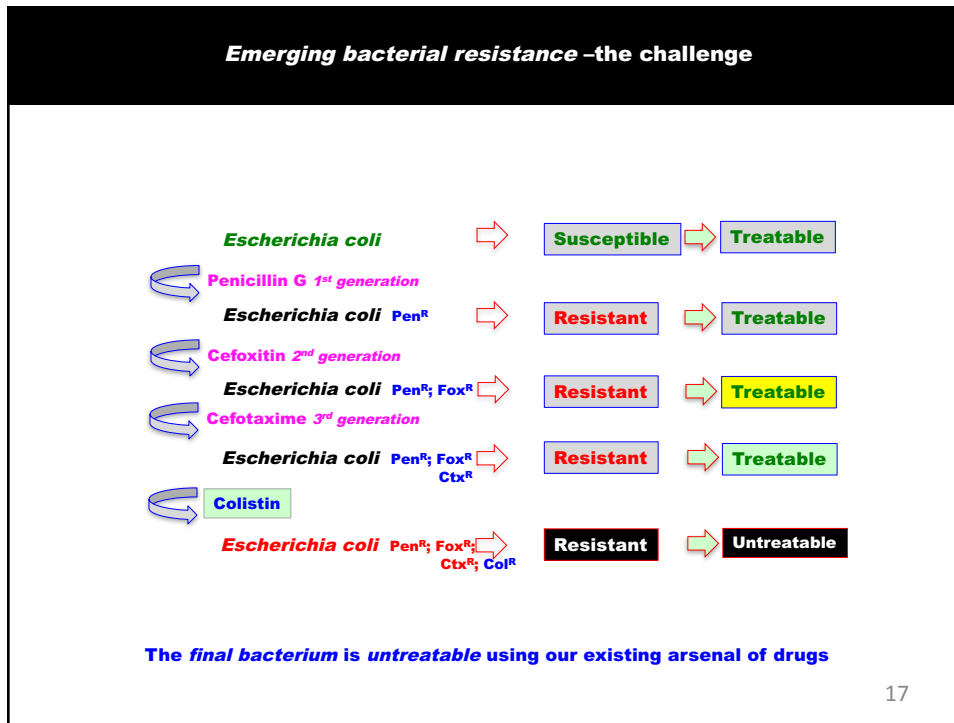


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Antimicrobial susceptibility testing (AST methods) -

Disc Diffusion

Disc code
AMP, ampicillin KF, cephalothin
C, chloramphenicol P, penicillin G
E, erythromycin SXT, trimethoprim-sulphamethoxazole
ENR, enrofloxacin TE, tetracycline
Numbers on discs indicate the drug content (µg); for penicillin the number indicates international units

Broth Microdilution

E-Test

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Global epidemiology of antibiotic resistance & use -

- decreasing efficacy of antibiotics used to treat infections has quickened in recent years with the **emergence of carbapenem-resistant Enterobacteriaceae**
- in high-income countries, high rates of antibiotic use has been documented in hospitals, the community and in agriculture
- **increased/continued selection pressure has sustained the persistence of resistant bacterial strains**
- **in developing countries, antibiotic resistance is also increasing**
- resistance emerges following mutations of genes on the chromosome and/or following the acquisition of a mobile genetic element
- these events gave rise to resistant strains including **MRSA; E. coli ST131; Klebsiella pneumoniae ST258 & Salmonella Typhimurium DT104**

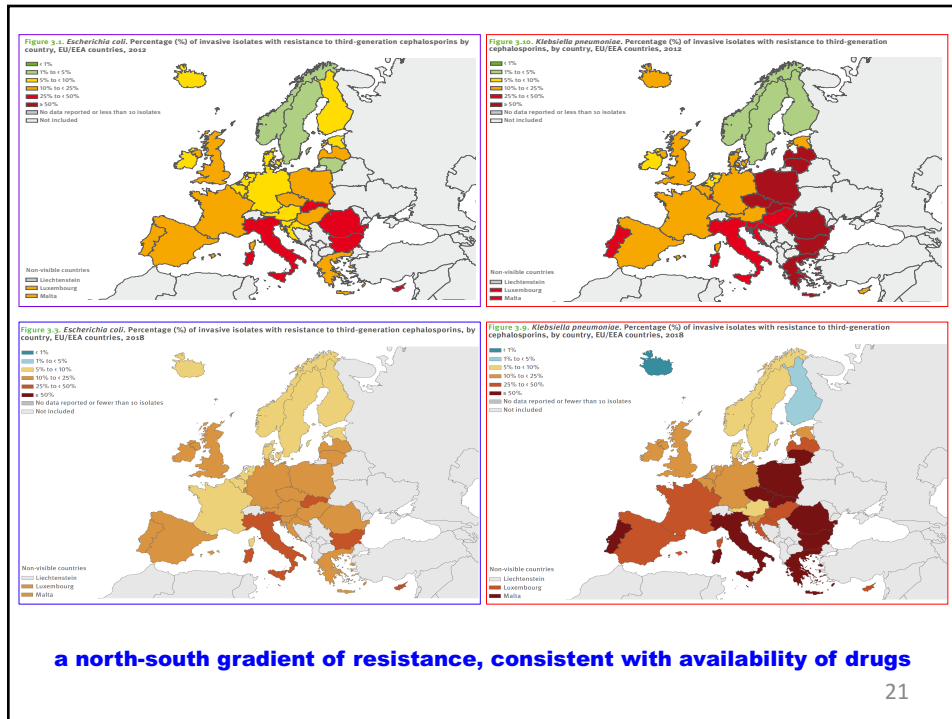
over the counter

20

Figure 1: Trends in retail sales of carbapenem antibiotics for Gram-negative bacteria. Based on data obtained from IMS Health's MIDAS™ database. *An IMS grouping of Benin, Burkina Faso, Cameroon, Congo (Brazzaville), Gabon, Guinea, Ivory Coast, Mali, Senegal, and Togo.

[Laxminarayan et al., The Lancet (2013) 13: 1057-1098]

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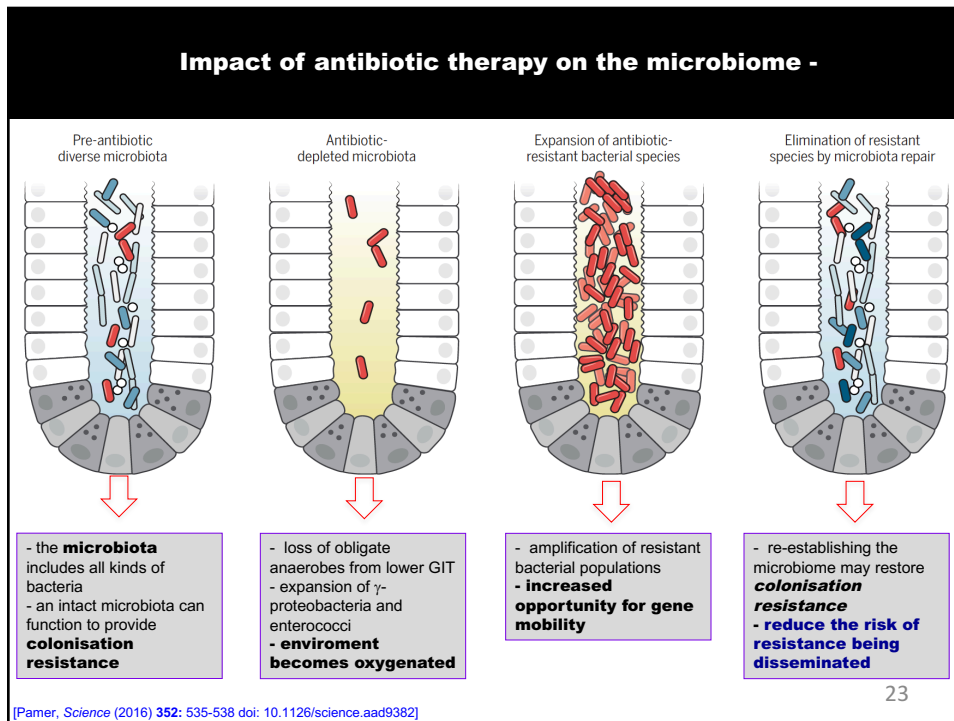
Antibiotic therapy can be likened to a four-edged sword -

- **Personal benefit** through individual treatment
- **Community benefit** by controlling the spread of disease
- **Emergence of resistant bacteria**, following selection during treatment and its impact on the community health
- **Costs associated with the animals/individuals health via collateral damage of the drug on bacteria that normally live on or in healthy animals/individuals- The Microbiome**



The Microbiome-

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Antimicrobial agents in a *One Health* context -

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Sales of antibacterial agents in 10 European states in 2007-

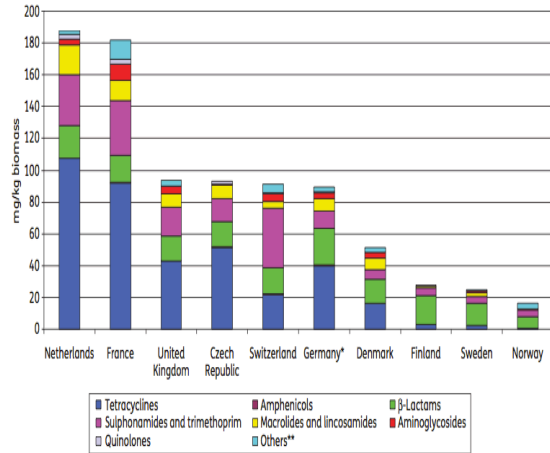


Figure 1. Amounts, in mg, of veterinary antibacterial agents sold in 2007 per kg biomass of pig meat, poultry meat and cattle meat produced plus estimated live weight of dairy cattle. *2005 data. **The substances included vary from country to country.

[Grave et al., J. Antimicrob. Chemother. (2010) 65: 2037-2040]

- data on annual sales of veterinary antibacterial compounds for therapeutic use are published annually by 10 European states

- **what treatments these drugs were used for is unclear**

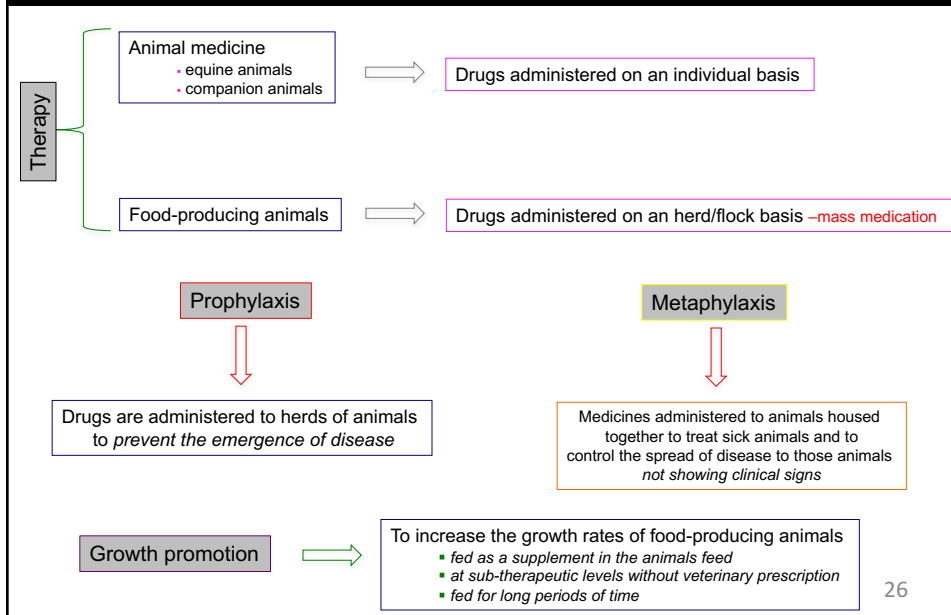
- amounts of veterinary antibacterial compound used relative to food-producing animal biomass varied from 18-188 mg/kg

- across the 10 states studied, **48% of sales were accounted for by tetracyclines**; 17% STX and 16% beta-lactams

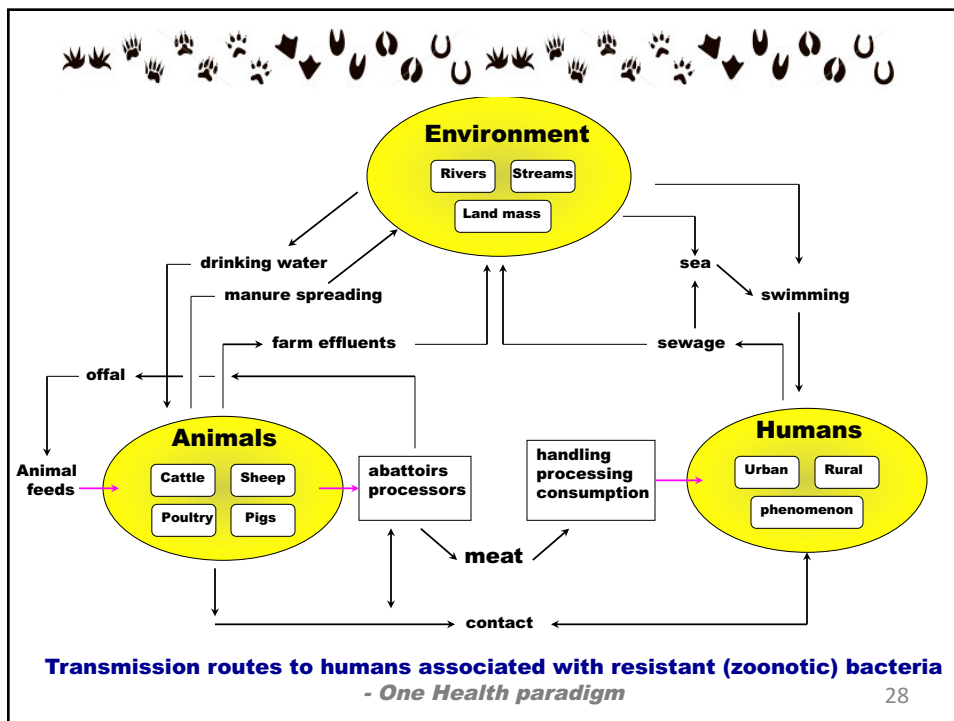
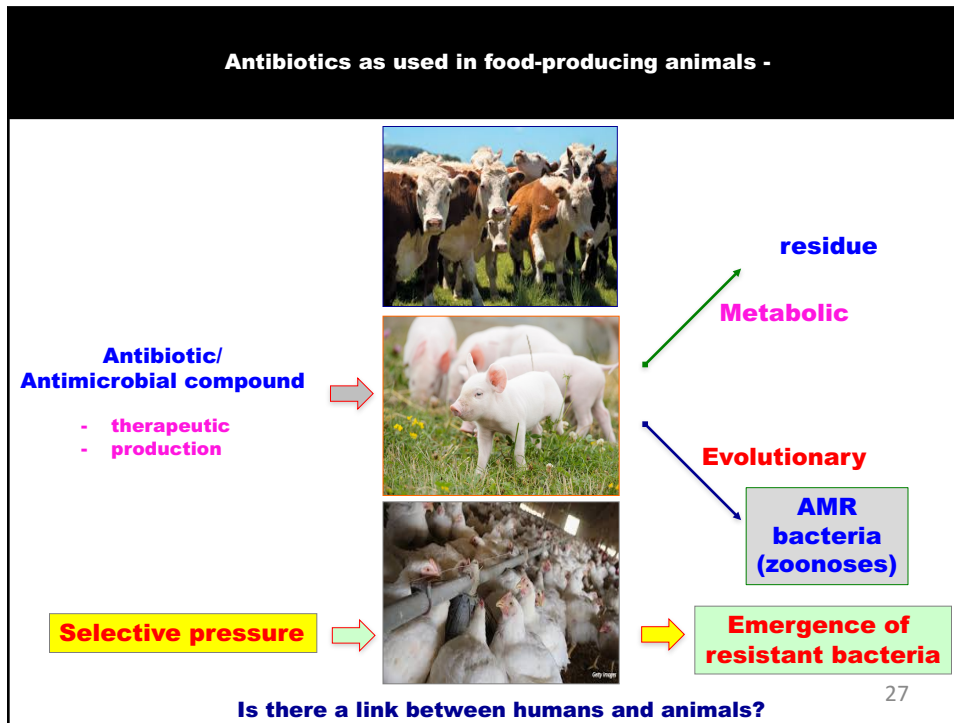
- **usage patterns varied between countries** (tetracyclines more frequently used in NL; CZ & F)

- **cephalosporin sales accounted for between 1 and 7%, depending on the country**

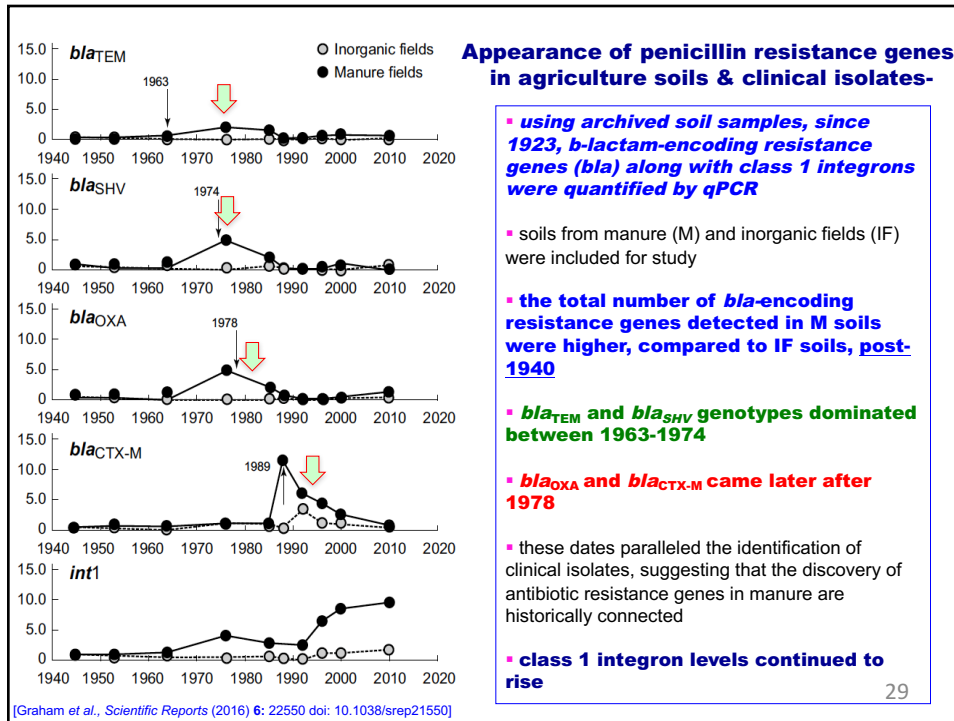
Antibacterial compound usage in animals -



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Illustrative examples of AMR transmission from animals to humans -

| | | |
|--|--|--|
| <p>Aminoglycosides</p> <p>Nourseothricin (NTC) <i>sat; stat; nat</i></p> | <p>Glycopeptides</p> <p>Avoparcin <i>vanA</i> Vancomycin-resistant enterococci (VRE)</p> | <p>Polymyxins</p> <p>Polymyxin E (colistin) <i>mcr-1; mcr-2; mcr 3-9</i> Colistin-resistant Enterobacteriaceae</p> |
|--|--|--|

Antimicrobial-based growth promoters contribute to resistance dissemination

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World Health Organisation (WHO) –important drug classes preserved for human health -

- **Critically important antibiotics (CIA)-**
 include the following- gentamycin; streptomycin; apramycin; neomycin;
 kanamycin; ampicillin; amoxicillin/clavulanic acid; carbapenems; 3rd/4th
 generation cephalosporins; monobactams; penicillin G; nalidixic acid;
 fluoroquinolones (FQ); macrolides; linezolid; **tigecycline**; **colistin**
- **Highly important antibiotics-**
 include the following- phenicols; 1st/2nd generation cephalosporins;
 cloxacillin; lincosamides; (virginiamycin); sulphonamides;
 chlorotetracycline; oxytetracycline
- All other antimicrobial compounds are classified as important-
- **the high priority classes-**
 include the following- 3rd/4th generation cephalosporins; FQ;
 macrolides & glycopeptides

carbapenems; oxazolidinones & **glycylcyclines** are not licensed for veterinary use

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Antimicrobial resistance is spreading worldwide -

- **almost all disease-causing bacteria resistant**
- b-Lactamase-mediated resistance gave rise to > 1,000 enzymes now known to inactivate b-lactam drugs, since before 1990 & some examples include-
 - **New Delhi metallo-b-lactamase-1 (NDM-1) discovered in 2008 and has now spread worldwide [environmental origin]**
 - **carbapenem-resistant *Klebsiella pneumoniae* (KPC) are commonly encountered in clinical settings**
 - **extended spectrum b-lactamase (ESBL)-encoding genes are rapidly spreading through Enterobacteriaceae from foods of animal origin and clinical settings**
- these trends are globally consistent

Risk factors associated with the dissemination of antibiotic resistance

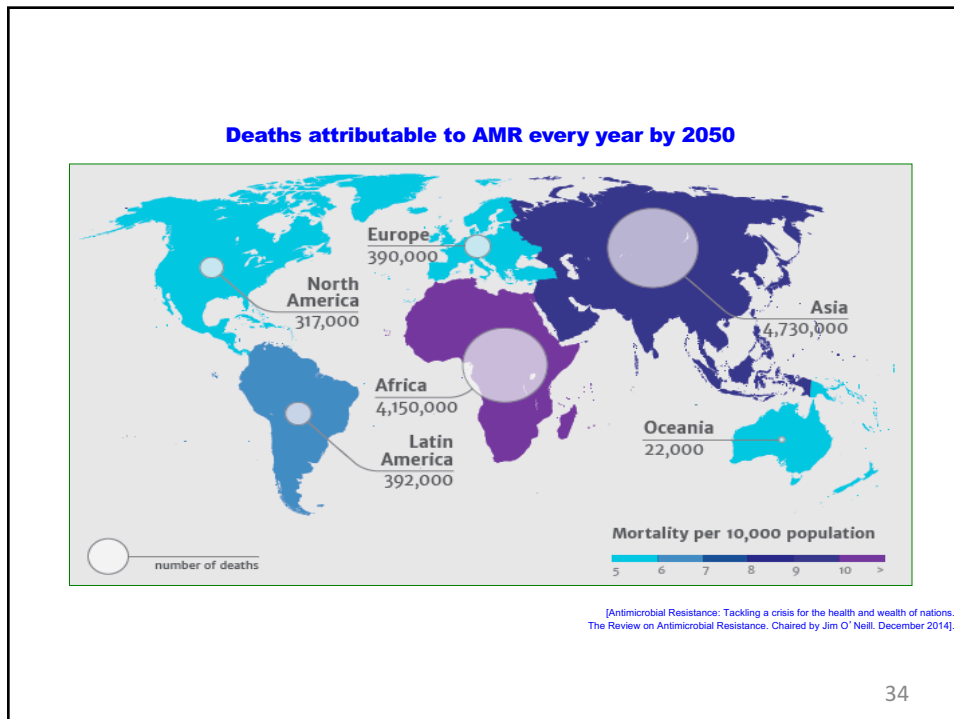
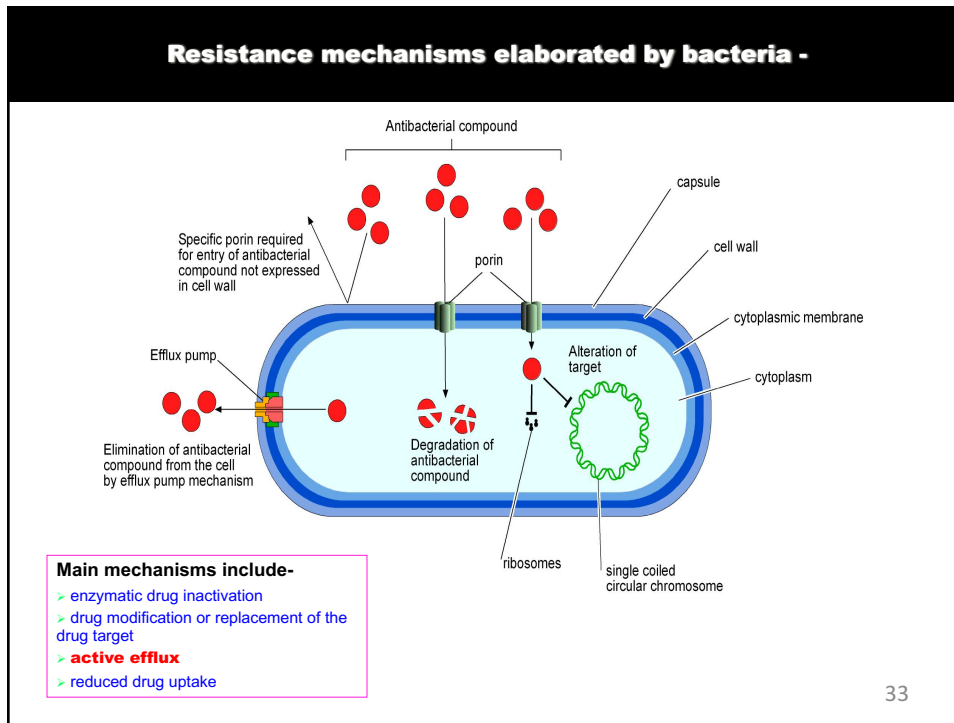


- bacterial interspecies gene transmission
- poor sanitation
- hygiene in hospitals & the community
- agriculture/veterinary use
- global trade (in food)
- foreign travel

the general public do not appear to understand the challenge facing society

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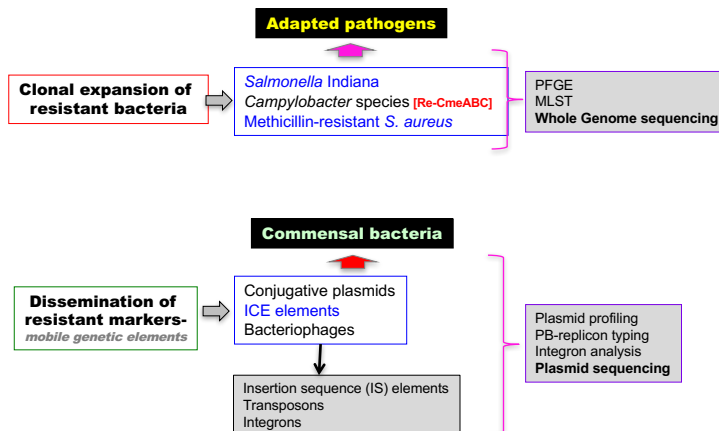
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How resistance to antibiotics/antimicrobial compounds becomes disseminated -

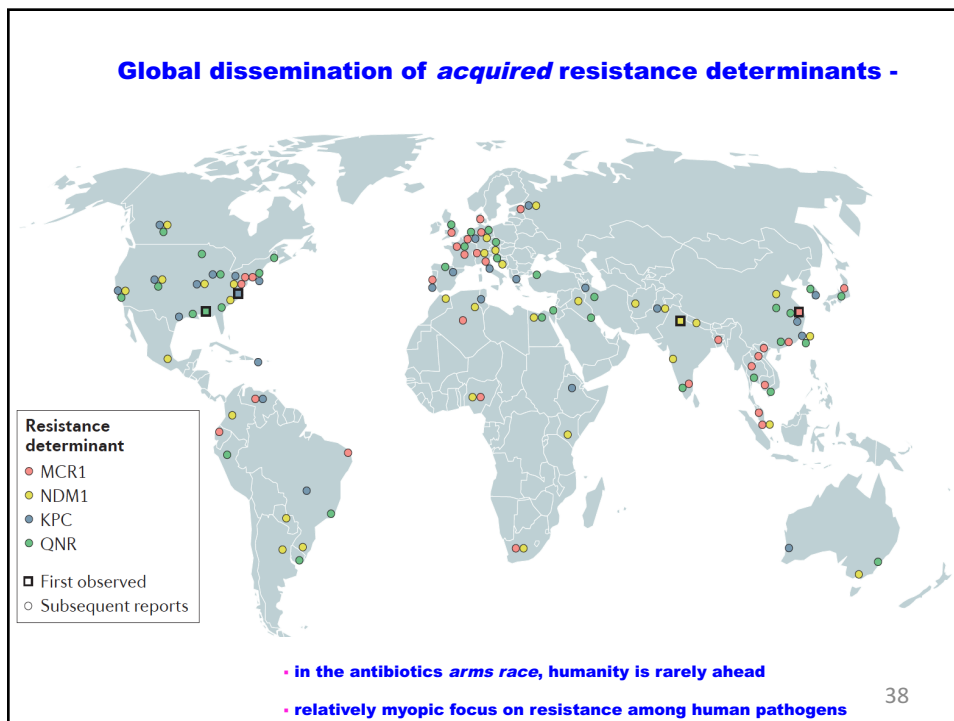
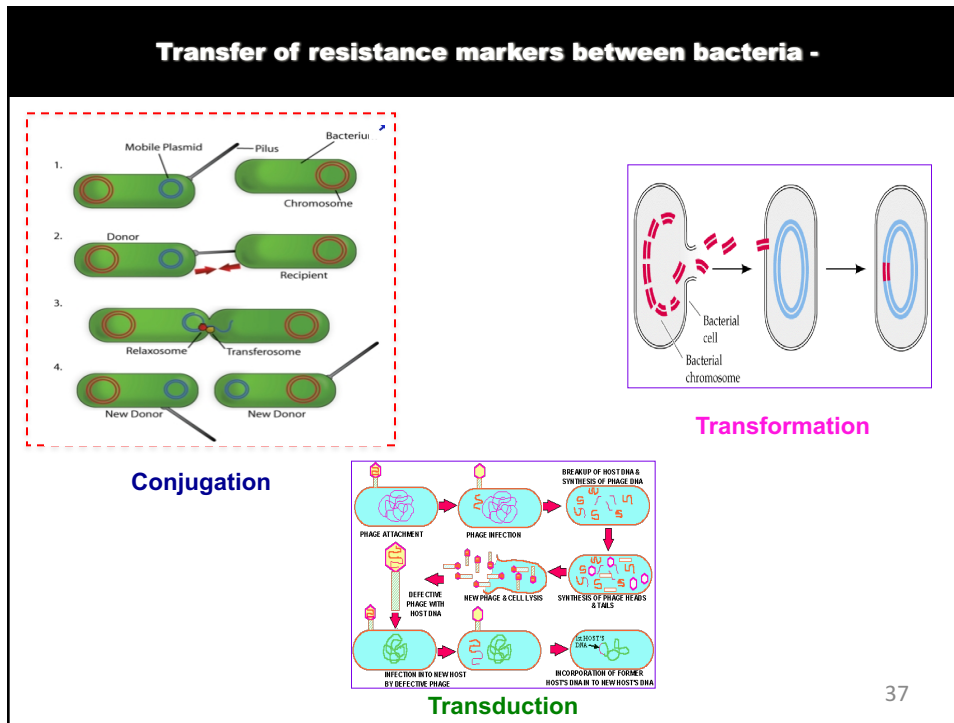
35

Emergence and expansion of adapted bacterial zoonoses -



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Examples of AMR genotypes-

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Salmonella recovered from whole chicken carcasses
at retail outlets -

Experimental strategy-

- conventional microbiological methods &
- AST testing

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Salmonella epidemiology in China -

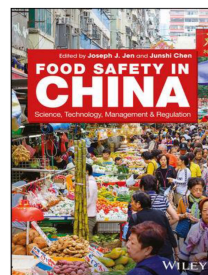
- *Salmonella*-contaminated chicken products are an important vehicle for much of the food-borne infection cases reported in China
- Some 40% of the 2,203 food-borne disease outbreaks reported between 2006-2010 in China were attributed to microorganisms
- **Of these 70-80% had a defined aetiology linked to *Salmonella***
- ***Salmonella* infections have become a major public health issue**
- Most cases of salmonellosis are self-limiting and do not require chemotherapeutic intervention
- **Emergence of a *Salmonella* Indiana co-resistant to ceftriaxone and ciprofloxacin are now being regularly reported**



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Summary of the study -

- Aim of the study was to describe the serovar diversity and antimicrobial resistance phenotypes identified in NTS isolates from retail chicken carcasses across 6 provinces in China
- **Samples tested numbering 1,438 whole chickens tested, provided 2,210 *Salmonella* isolates for study**
- Serotyping was performed by conventional- and molecular-based methods and susceptibilities tested against a panel of 11 compounds representing nine classes of antimicrobial drug



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AST analysis of *Salmonella* serovars -

| Number of Class of Antibiotic Resistance | No. of strains of dominant serovars (Percentage) | | | | | | | | | | | |
|--|--|-----------------------|-------------------|--------------------|-----------------------|-----------------|----------------|-----------------|----------------|-------------------|----------------|--------------------------|
| | Total (n = 2210) | Enteritidis (n = 673) | Indiana (n = 365) | Infantis (n = 211) | Typhimurium (n = 163) | Agona (n = 162) | Derby (n = 81) | Rissen (n = 68) | Dabou (n = 58) | Thompson (n = 50) | Hadar (n = 46) | Other Serovars (n = 333) |
| 0 | 438 (19.8%) | 68 (10.1%) | 16 (1.6%) | 16 (7.6%) | 24 (14.7%) | 135 (83.3%) | 7 (8.6%) | 12 (17.6%) | 4 (6.9%) | 19 (38.0%) | 0 (0.0%) | 147 (44.1%) |
| 1 | 645 (29.2%) | 271 (40.3%) | 12 (3.3%) | 141 (66.8%) | 34 (20.9%) | 9 (5.6%) | 15 (18.5%) | 32 (47.1%) | 43 (74.1%) | 11 (22.0%) | 0 (0.0%) | 77 (23.1%) |
| 2 | 160 (7.2%) | 20 (3.0%) | 19 (5.2%) | 15 (7.1%) | 9 (5.5%) | 0 (0%) | 10 (12.3%) | 3 (4.4%) | 11 (19.0%) | 0 (0.0%) | 42 (91.3%) | 31 (9.3%) |
| 3 | 175 (7.9%) | 128 (19.0%) | 18 (4.9%) | 1 (0.5%) | 10 (6.1%) | 0 (0%) | 4 (4.9%) | 2 (2.9%) | 0 (0.0%) | 0 (0.0%) | 2 (4.3%) | 10 (3.0%) |
| 4 | 198 (9.0%) | 148 (22.0%) | 15 (4.1%) | 0 (0%) | 16 (9.8%) | 1 (0.6%) | 9 (11.1%) | 2 (2.9%) | 0 (0.0%) | 0 (0.0%) | 2 (4.3%) | 5 (1.5%) |
| 5 | 101 (4.6%) | 14 (2.1%) | 25 (6.9%) | 1 (0.5%) | 19 (11.7%) | 6 (3.7%) | 9 (11.1%) | 7 (10.3%) | 0 (0.0%) | 1 (2.0%) | 0 (0.0%) | 19 (5.7%) |
| 6 | 209 (9.4%) | 21 (3.1%) | 62 (17.0%) | 37 (17.5%) | 27 (16.6%) | 11 (6.8%) | 3 (3.7%) | 9 (13.2%) | 0 (0.0%) | 6 (12.0%) | 0 (0.0%) | 33 (9.9%) |
| 7 | 184 (8.3%) | 3 (0.5%) | 109 (29.9%) | 0 (0%) | 24 (14.7%) | 0 (0%) | 24 (29.6%) | 1 (1.5%) | 0 (0.0%) | 13 (26.0%) | 0 (0.0%) | 10 (3.0%) |
| 8 | 100 (4.5%) | 0 (0%) | 99 (27.1%) | 1 (0.5%) | 0 (0%) | 0 (0%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 0 (0.0%) | 1 (0.3%) |
| ≥3 (MDR) | 967 (43.8%) | 314 (46.7%) | 328 (89.9%) | 39 (18.5%) | 96 (58.9%) | 18 (11.1%) | 49 (60.5%) | 21 (30.9%) | 0 (0.0%) | 20 (40.0%) | 4 (8.7%) | 78 (23.4%) |
| ≥5 | 594 (26.9%) | 38 (5.7%) | 295 (80.8%) | 38 (18.0%) | 70 (42.9%) | 17 (10.5%) | 36 (44.4%) | 17 (25.0%) | 0 (0.0%) | 20 (40.0%) | 0 (0.0%) | 63 (18.9%) |

AST summary –

- some 438 isolates tested were found to be susceptible;
- 1,772 were resistant to at least one antimicrobial compound;
- resistance to nalidixic acid was common (70% of the collection) followed by resistance to ampicillin (43%), tetracycline (42%);
- none of the isolates were resistant to carbapenems;
- 55% of the collection were defined as having a MDR phenotype (3 or more classes)

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[Hu, et al., (2017) Food Control 81: 46-54]

| Antimicrobial resistance profile | Number of Isolates | Number of ESBLs positive isolates |
|---|--------------------|-----------------------------------|
| CHL-CIP-NAL-AMP-CAZ-CTX-SXT | 1 | 1 |
| CHL-CIP-NAL-AMP-SAM-CAZ-CTX-SXT | 1 | 0 |
| CHL-CIP-NAL-AMP-SAM-TET-CTX-SXT | 13 | 13 |
| CIP-NAL-AMP-SAM-CTX | 1 | 1 |
| CIP-NAL-AMP-TET-CTX | 1 | 1 |
| GEN-CHL-CIP-NAL-AMP-CTX-SXT | 2 | 2 |
| GEN-CHL-CIP-NAL-AMP-SAM-CAZ-CTX-SXT | 15 | 14 |
| GEN-CHL-CIP-NAL-AMP-SAM-CTX | 4 | 4 |
| GEN-CHL-CIP-NAL-AMP-SAM-CTX-SXT | 19 | 18 |
| GEN-CHL-CIP-NAL-AMP-SAM-TET-CAZ-CTX | 11 | 11 |
| GEN-CHL-CIP-NAL-AMP-SAM-TET-CAZ-CTX-SXT | 32 | 32 |
| GEN-CHL-CIP-NAL-AMP-SAM-TET-CTX | 9 | 9 |
| GEN-CHL-CIP-NAL-AMP-SAM-TET-CTX-SXT | 66 | 65 |
| GEN-CHL-CIP-NAL-AMP-TET-CTX | 1 | 1 |
| GEN-CHL-CIP-NAL-AMP-TET-CTX-SXT | 1 | 1 |
| GEN-CIP-NAL-AMP-CTX-SXT | 1 | 1 |
| GEN-CIP-NAL-AMP-SAM-CTX-SXT | 1 | 1 |
| GEN-CIP-NAL-AMP-SAM-TET-CTX | 1 | 1 |
| GEN-CIP-NAL-AMP-SAM-TET-CTX-SXT | 3 | 3 |
| Total | 183 | 179 |

***Salmonella* Indiana
ESBL & FQ resistant profiles-**

- *Salmonella* Indiana were the most resistant of the serovars detected with 98% being resistant to all compounds tested **with the exception of carbapenems**
- *Salmonella* Indiana was also found to be resistant to (fluoro)quinolones
- 183 *S. Indiana* isolates were co-resistant to ciprofloxacin and cefotaxime of which 179 were ESBL-positive
- more than half of the isolates represented by this serovar were found to be resistant to nine or more antimicrobial compounds

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**Salmonella harbouring *mcr-1* gene
isolated from food in China between 2011-2016-**

Experimental strategy-

- o conventional microbiological methods;
- o AST testing;
- o WGS (SMRT® platform) &
- o bioinformatic analyses

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Colistin an old drug that has become important (again) -

* *colistin is a cationic peptide that belongs to the family of polymyxins*

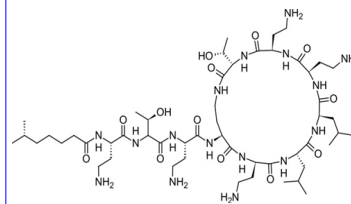
* *discovered in 1947 and has a complex biochemical structure*

* *fell into disuse, by 1980's due to nephrotoxicity*

* *these resistance mechanisms are thought to be mediated by genes located on the bacterial chromosome **NOT TRANSFERRABLE TO OTHER BACTERIA***

* *in 2015, a transferrable mechanism denoted as *mcr-1* was described, located on a plasmid that moved at high frequency*

* **mcr-2* has just been in June, 2016, in Belgium and associated with bacterial isolates cultured from bovine and porcine animals*



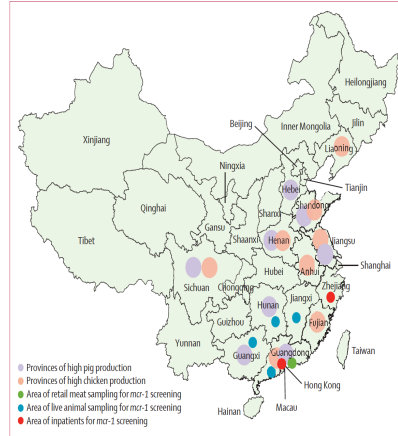
Polymyxin E (Colistin)-

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[Liu et al., (2015) *Lancet Infect. Dis.*16: 161-168. doi: 10.1061/S1473-3099(15)00424-7]

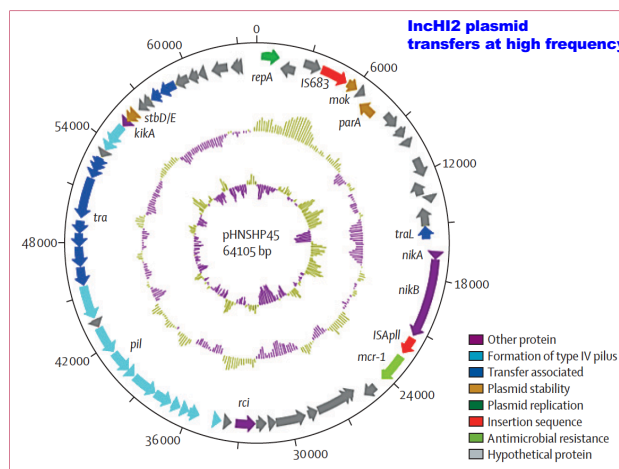
- **China is the worlds biggest poultry and pig producer** [in 2014, 17.5- and 56.7-million tonnes respectively]
- **China was one of the largest users of colistin for agricultural production** [global market value in veterinary drugs increased from \$ 8.7 billion in 1992 to \$ 20.1 billion in 2010 *and this is expected to grow to \$ 43 billion by 2018*].
- **Global demand for colistin in agriculture is expected to reach 16,500 tonnes by 2021**



Geographical locations in China positive for colistin-resistant *E. coli*

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Transferrable colistin resistance –the *mcr-1* gene



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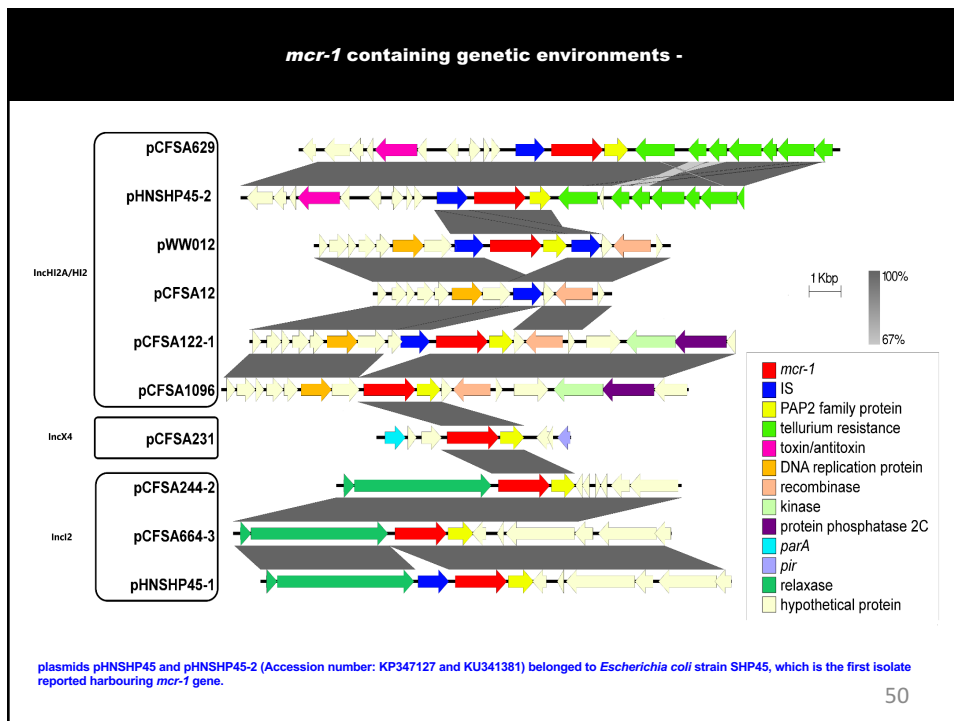
[Liu et al., (2015) *Lancet Infect. Dis.*16: 161-168. doi: 10.1061/S1473-3099(15)00424-7]

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Seven colistin isolates of 2,588 *Salmonella* species cultured from foods in China–

| Bacterial isolate | CFSA664 | CFSA122 | CFSA12 | CFSA244 | CFSA1096 | CFSA231 | CFSA629 |
|--|----------------|---|--|----------------|--|-----------------------|--|
| Serotype | Indiana | Typhimurium | Typhimurium | Typhimurium | London | Derby | Typhimurium |
| MLST type | 17 | 34 | 34 | 34 | 155 | 40 | 34 |
| Year.month | 2011.6 | 2013.5 | 2014.5 | 2014.4 | 2015.10 | 2016.10 | 2016.7 |
| Region | Jiangsu | Guangxi | Guangxi | Jiangxi | Hubei | Hubei | Guangdong |
| Food origin | Retail chicken | Dumpling (Retail) | Retail Pork | Retail Pork | Retail Pork | Dumpling (Restaurant) | Egg |
| Resistant antimicrobial classes | 8 | 7 | 4 | 4 | 10 | 7 | 6 |
| ESBLs | + | - | - | - | + | - | + |
| Number of plasmid | 3 | 2 | 1 | 2 | 1 | 1 | 1 |
| Replicon type and size of plasmids harbouring mcr-1 | Incl2 (61 kbp) | IncH2A/HI2 (182 kbp) | IncH2A/HI2 (148 kbp) | Incl2 (60 kbp) | IncH2A/HI2 (297 kbp) | IncX4 (33 kbp) | IncH2A/HI2 (211 kbp) |
| Antimicrobial resistance genes on plasmids harbouring mcr-1 | <i>mcr-1</i> | <i>mcr-1</i> , <i>aph(4)-Ia</i> , <i>aac(3)-Iva</i> , <i>aph(3)-Ia</i> , <i>aadA1</i> , <i>aadA2</i> , <i>bla_{CTX-M-15}</i> , <i>asc(6)-Ib-cr</i> , <i>floR</i> , <i>catB4</i> , <i>cmlA1</i> , <i>arr-3</i> , <i>sul2</i> , <i>sul1</i> , <i>sul3</i> | <i>aadA2</i> , <i>aadA1</i> , <i>cmlA1</i> , <i>sul3</i> , <i>dfrA12</i> , <i>Δmcr-1</i> | <i>mcr-1</i> | <i>mcr-1</i> , <i>aph(3)-Ia</i> , <i>aadA22</i> , <i>aph(6)-Id</i> , <i>aac(3)-IId</i> , <i>bla_{TEM-1B}</i> , <i>bla_{CTX-M-55}</i> , <i>qnrS1</i> , <i>inu(I)</i> , <i>mph(A)</i> , <i>floR</i> , <i>arr-2</i> , <i>sul3</i> , <i>tet(A)</i> , <i>dfrA14</i> | <i>mcr-1</i> | <i>mcr-1</i> , <i>aac(3)-Iva</i> , <i>aph(4)-Ia</i> , <i>bla_{CTX-M-14}</i> , <i>fosA3</i> |

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Selection of plasmids co-expressing resistance to tigecycline and florphenicol -

Experimental strategy-

- o conventional microbiological methods;
- o AST testing;
- o WGS (SMRT® platform) &
- o bioinformatic analyses

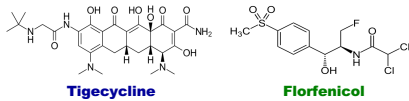
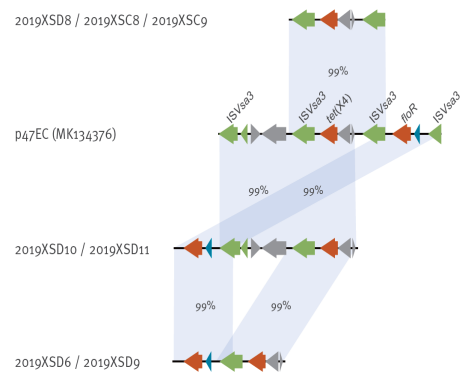
▪ *Escherichia coli* were cultured from food-producing animals

▪ bacteria were isolated following selection on tigecycline & seven expressed a MIC (8-32 mg/L) above the breakpoint

▪ the *tet(X4)* gene was identified and contained within a gene cassette which may support its dissemination by horizontal means on an *IncFII* plasmid

▪ WGS analysis of these isolates identified ST101 & ST48

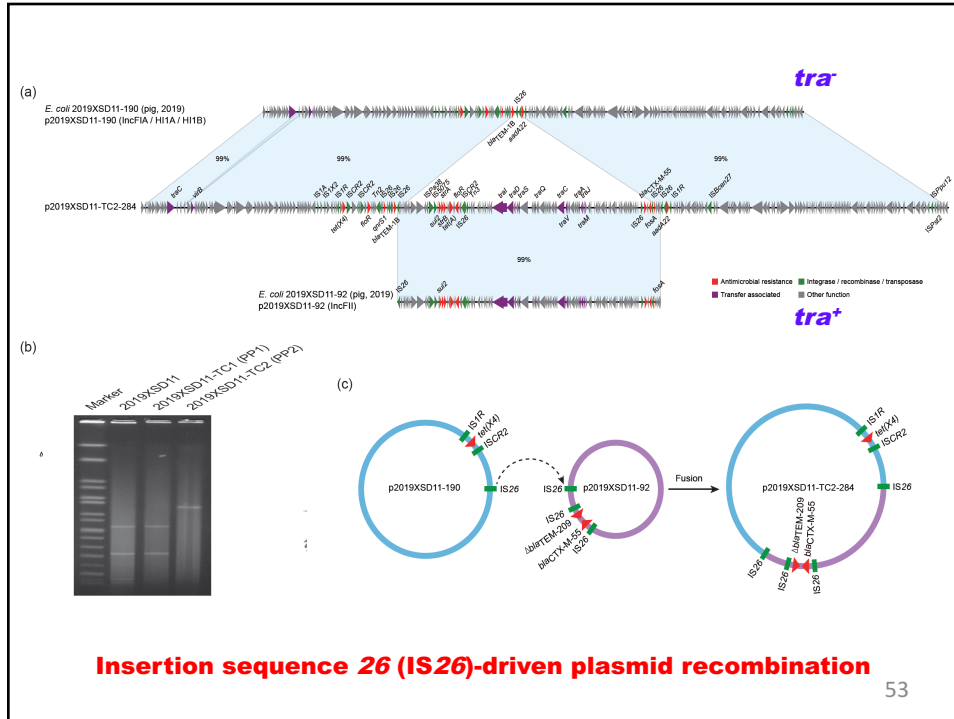
▪ the *floR*-encoding gene was flanked by *IS6100*, suggesting that tigecycline and florphenicol resistance are being co-selected



[Li et al., Euro. Surveill. (2019) 24: pii=19003]
 [He et al., Nat. Microbiol. (2019) 10.1038/s41564-019-0445-2]

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Metagenomics of antimicrobial resistance -

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Metagenomics to characterize microbial ecology -

Metagenomics can be applied to study the bacterial composition of samples recovered directly from any environment -

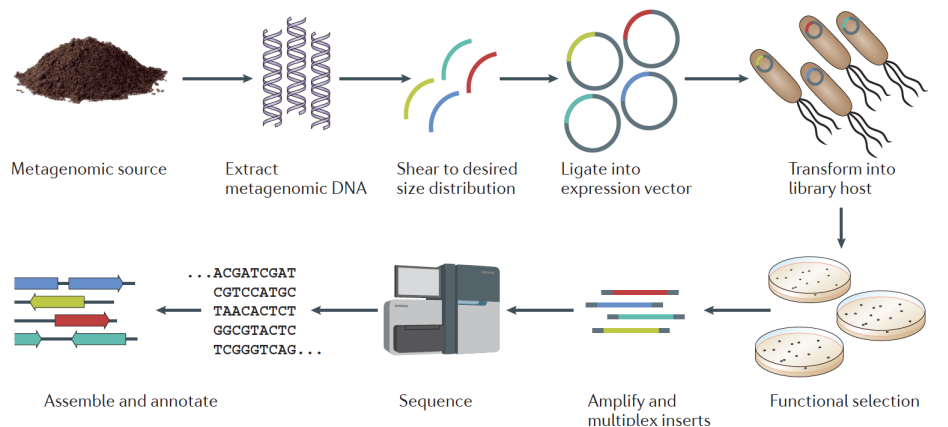
- **Targeted (PCR-based) metagenomics;** PCR used to detect various bacterial-encoded 16S rDNA genes and this can be applied as a semi-quantitative method

- **Functional metagenomics;** involves the cloning of a DNA fragment into a heterologous genetic background and selecting for a defined phenotype using a selective culture medium, followed by subsequent DNA sequencing to characterise any positive clone

- **Sequence-based metagenomics;** DNA from an environment of interest is purified; fragmented and size-separated then sequenced directly; sequences obtained are compared to entries in the current databases

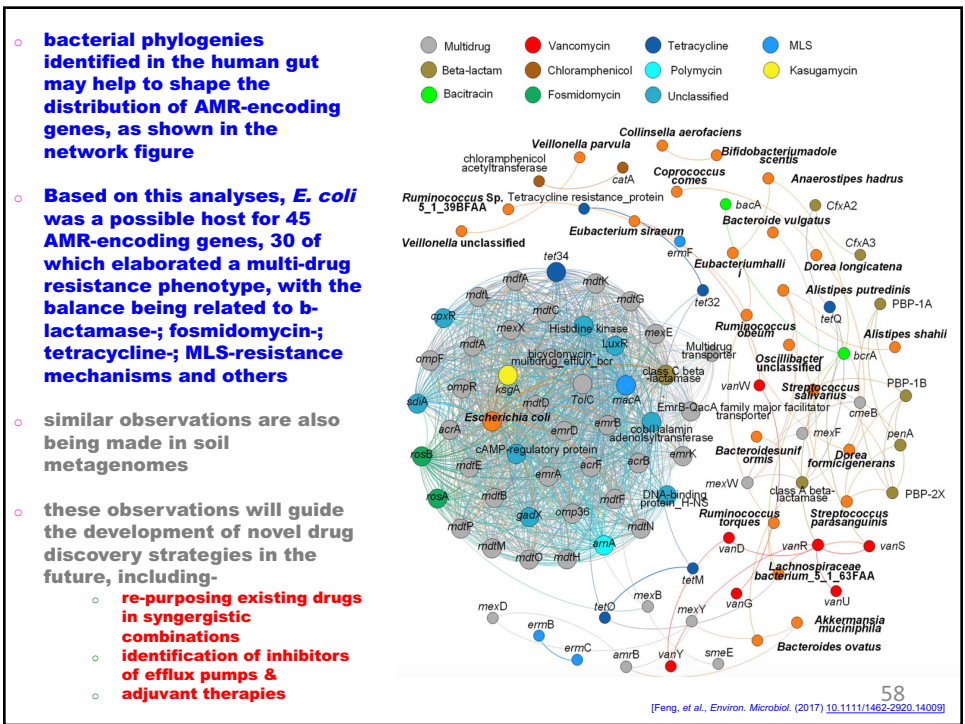
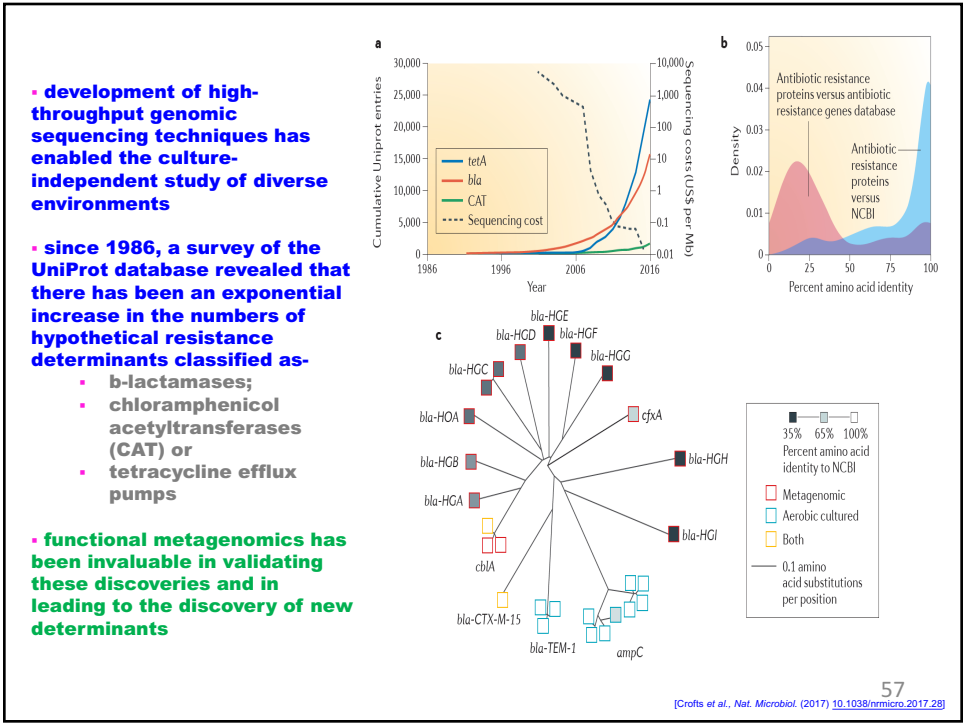
55
 [Penders et al., (2013) *Frontiers in Microbiol*, doi: 10.3389/fmicb.2013.00087]

Steps involved in functional metagenomics -



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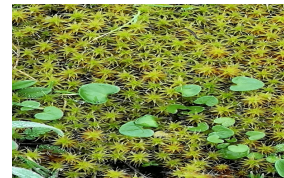
Conclusions -

- AMR is a *One Health* challenge;
- AMR resistance-encoding genes can be disseminated widely by several mechanisms;
- *matagenomic libraries may be helpful in fueling the discovery of novel antimicrobial compounds*

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Thank you



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| www.webbertraining.com/schedulep1.php | |
|--|--|
| February 19, 2020 | <p><i>(South Pacific Teleclass)</i> DEVELOPING AND IMPLEMENTING A PERSONAL PROTECTIVE EQUIPMENT TRAINING PROGRAMME FOR HIGH-CONSEQUENCE INFECTIOUS DISEASE PREPAREDNESS Speaker: Ruth Barratt, University of Sydney, Faculty of Medicine</p> |
| February 27, 2020 | <p>ANTIBIOTIC STEWARDSHIP IN NURSING HOMES Speaker: Prof. Patricia Stone, Columbia University, School of Nursing</p> |
| March 3, 2020 | <p><i>(European Teleclass)</i> THE EFFICACY OF INFECTION PREVENTION AND CONTROL COMMITTEES IN AFRICAN SETTINGS Speaker: Eltony Mugomeri, Africa University, Zimbabwe</p> |
| March 12, 2020 | <p><i>(FREE Teleclass)</i> THE BUZZ AROUND MOSQUITOES AND MOSQUITO-BORNE DISEASES Speaker: Dr. Marcia Anderson, Environmental Protection Agency</p> |
| March 19, 2020 | <p>INFECTION PREVENTION AND CONTROL IN HOME CARE AND HOSPICE: COMMON COMPLIANCE ISSUES Speaker: Mary McGoldrick, Home Health Systems, Inc.</p> |
| April 16, 2020 | <p>WATERBORNE PATHOGENS: WHY IS THEIR PROFILE CHANGING? Speaker: Prof. Syed A Sattar, Centre for Research on Environmental Microbiology, Canada</p> |

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